### STERIC COURSE OF THE HISTIDASE REACTION

# J.RÉTEY, H.FIERZ, W.P.ZEYLEMAKER

Organisch-chemisches Laboratorium, Eidgenössische Technische Hochschule, Universitätstrasse 6, 8006 Zürich and Laboratory of Biochemistry, B.C.P. Jansen Institute,
University of Amsterdam, Plantage Muidergracht 12, Amsterdam, The Netherlands

Received 18 December 1969

#### 1. Introduction

Histidase (histidine ammonia lyase, E.C. 4.3.1.3) catalyses the practically irreversible\* conversion of L-histidine to urocanic acid and ammonia according to equ. (1). In the course of the reaction one of the heterotopic [1] hydrogen atoms at C-3 of L-histidine is

slowly exchanged with the protons of water and this hydrogen is lost in the elimination reaction [2]. It will subsequently be shown that the  $H_R$  atom [3] of C-3 is involved both in the exchange and in the overall reaction and hence the elimination of ammonia from L-histidine occurs in a "trans" manner.

#### 2. Methods

Enzymically labelled L- $^3$ H-histidine was prepared essentially as described in [2]. A few  $\mu$ moles of the tritiated histidine were freeze-dried and diluted with 1.5 g of unlabelled L-histidine hydrochloride. After chromatography on Dowex-50 (eluent 4 N HBr) 2 g

\* Williams and Hiroms [11] recently showed that on very long incubation histidase catalyses the conversion of urocanate and ammonia to L-histidine.

crystalline L-histidine dihydrobromide were obtained (2280 cpm/mmole), the radioactivity of which remained unchanged on repeated chromatography.

2.1. Degradation of  $L^{-3}H$ -histidine to succinic acid

The following procedure was used: 580 mg L-3Hhistidine dihydrobromide were dissolved in 4 ml water and 0.87 ml HBr (48%). A solution of 125 mg NaNO<sub>2</sub> in 1.5 ml water was slowly added at 0°C and the reaction mixture stirred at 20°C for 2 hr. After addition of 100 mg PtO2, the mixture was hydrogenated at room temperature until the uptake of H2 ceased. A total of 43 ml H<sub>2</sub> were consumed. The catalyst was subsequently removed and the solution adjusted to pH 7 with 1 N KOH. A saturated aqueous solution of 1 g KMnO<sub>4</sub> was added dropwise to the mixture at 60°C. After 1½ hr stirring another portion of 1 g KMnO<sub>4</sub> was added and the reaction allowed to proceed for a further 1½ hr. In order to reduce MnVII and MnIV to  $\mathrm{Mn^{II}}$ , 15 ml of 38%  $\mathrm{NaHSO_3}$  were introduced to the solution. After acidification with 2 N H<sub>2</sub>SO<sub>4</sub>, the succinic acid was extracted continuosly with ether and purified by chromatography on Dowex-1 and several recrystallisations from water as described in [4]. The yield after 3 recrystallisations was 31 mg. The same degradation procedure with unlabelled L-histidine dihydrobromide, and using <sup>3</sup>H-H<sub>2</sub>O as solvent, resulted in practically no incorporation of tritium into the succinic acid.

2.2. Determination of chirality of <sup>3</sup>H-succinic acid

The chirality determination was carried out as described in [5]. Reference (R)-, (S)- and (RS)-<sup>3</sup>H-succinate samples were oxidised to 70% conversion with the same preparation of soluble succinate dehydrogen-

ase. Succinic and fumaric acid samples were separated and purified as described in [4]. The results of the radioactivity measurements are summarised in the table.

#### 3. Results and discussion

A detailed study of the mechanism of the histidase reaction has revealed [2] that in addition to the overall conversion of L-histidine to urocanic acid, the following two partial reactions are catalysed by the enzyme; i) dismutation bettween L-histidine and <sup>14</sup>Curocanic acid and ii) exchange of hydrogen atoms between L-histidine and water. The tritium atom introduced to L-histidine by the latter process was localised in the methylene group by appropriate degradation procedures [2]. Furthermore, enzymically tritiated Lhistidine was converted by the enzyme into tritium free urocanic acid. This implies that only one of the diastereotopic H atoms at C-3 of L-histidine is exchangeable and that the same hydrogen is lost in the elimination reaction. In the present work, it is shown that only the H<sub>R</sub> atom of L-histidine (equ. 1) is reactive both in the exchange and in the overall reaction. This is achieved by reductive elimination of the amino group of enzymically tritiated L-histidine and subsequent degradation of the imidazolyl propionic acid to <sup>3</sup>H-succinate (equ. 2). The latter is shown to possess the (R)-configuration by a method [5] based on the

difference in isotope effects for the enzymic removal of tritium from positions corresponding to  $H_R$  and  $H_S$  in succinate (see formula 1). Thus on partial oxida-

tion of the chiral <sup>3</sup>H-succinic acids by succinate dehydrogenase the enrichment of tritium in the starting material is faster when the substrate has the (R)-configuration than when it has the (S)-configuration (cf. table). In spite of recent work [2, 6] the precise mechanism of the histidase reaction is still unknown. The present results throw light on its stereochemistry by showing that it is a trans elimination process. Other enzymecatalysed reactions involving the elimination of ammonia, e.g. those catalysed by aspartase [7] and

Table

Substrate	Radioactivity				
	Starting succinate cpm/mmole	Succinate recovered		Fumarate produced	
		cpm/mmole	ratio compared with substrate (%)	cpm/mmole	ratio compared with substrate (%)
<sup>3</sup> H-succinate from L- <sup>3</sup> H-histidine	945 ± 30	1630 ± 50	173 ± 10	574 ± 15	58 ± 5
(R)- <sup>3</sup> H-succinate (reference)	1506 ± 15	2535 ± 15	168 ± 8	785 ± 15	52 ± 5
(S)-3H-succinate (reference)	1663 ± 15	2087 ± 15	125 ± 5	932 ± 10	56 ± 5
(RS)-3H-succinate (reference)	1581 ± 15	2321 ± 15	147 ± 7	922 ± 10	58 ± 5

 $\beta$ -methylaspartase [8-10], follow the same steric course although they differ from the histidase reaction in other respects.

# Acknowledgement

The technical assistence of Mr. B.Vogt is gratefully acknowledged.

## Addendum

After the preparation of this manuscript we received a preprint of a paper by I.L.Givot, T.A.Smith and R.H. Abeles in which they also show that the histidase reaction involves a "trans" elimination.

#### References

[1] K.Mislow and M.Raban, Topics in stereochemistry, Vol. 1, eds. Allinger and Eliel (1967) p.1.

- [2] A.Peterkofsky, J. Biol. Chem. 237 (1962) 787.
- [3] K.R.Hanson, J. Am. Chem. Soc. 88 (1966) 2731.
- [4] J.Rétey, J.Seibl, D.Arigoni, J.W.Cornforth, G.Ryback, W.P.Zeylemaker and C.Veeger, Nature 216 (1967) 1320.
- [5] W.P.Zeylemaker, C.Veeger, F.Kunz, J.Rétey and D.Arigoni, Chimia (in press).
- [6] T.A.Smith, F.H.Cordelle and R.H.Abeles, Arch. Biochem. Biophys. 120 (1967) 724.
- [7] O.Gawron and T.P.Fondy, J. Am. Chem. Soc. 81 (1959) 6333.
- [8] H.A.Barker, R.D.Smyth, E.J.Wawszkiewicz, M.N.Lee and R.M.Wilson, Arch. Biochem. Biophys. 78 (1958) 468.
- [9] H.J.Bright, L.L.Ingraham and R.E.Lundin, Biochim. Biophys. Acta 81 (1964) 576.
- [10] M.Sprecher and D.B.Sprinson, J. Biol. Chem. 241 (1966) 868.
- [11] V.R.Williams and J.M.Hiroms, Biochim. Biophys. Acta 139 (1967) 214.